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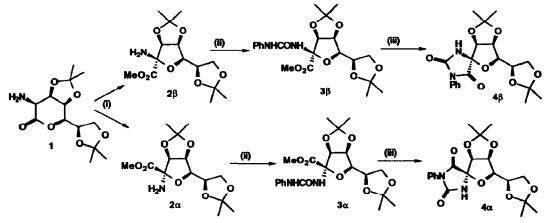
TRI- AND TETRA-PEPTIDES INCORPORATING AN α -Amino Acid at the Anomeric Position of Mannofuranose

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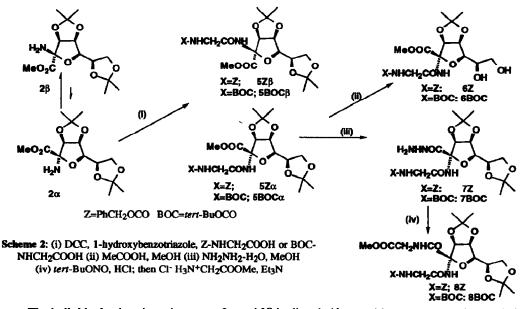
Abstract: The first examples of peptides containing an α -amino acid residue in which the α carbon is also the anomeric position of a carbohydrate are described.

The bromine-induced oxidative ring contraction of the aminolactone 1 to give the epimeric amino esters 2β and 2α in 60% and 17% yields, respectively, was the key step in the synthesis of some mannofuranose spirohydantoins¹ as analogues of the naturally occurring plant growth regulator, hydantocidin.² Although 2α and 2β slowly equilibrate in solution, reaction of the β anomer with phenyl isocyanate [Scheme 1] gave the urea 3β which cyclised in methanol to give the protected hydantoin 4β ; the minor α epimer on similar treatment gave 3α which cyclised to the epimeric hydantoin 4α , again without any apparent epimerisation. The reaction of the amines with phenyl isocyanate was faster than equilibration of 2α and 2β ; also, it was clear that equilibration of the epimeric ureas 3 and hydantoins 4 was negligible other than under highly acidic or basic conditions.



Scheme 1: (i) Br₂, NaOAc, MeOH; then Et₃N (ii) PhNCO, THF (iii) MeOH

The chemical stability of 3 and 4, together with the stereointegrity of the different anomers, indicated that it might be possible to incorporate amino acids at the anomeric position of sugars, such as 2, as components of peptides. Such amino acids would be of interest, *inter alia*, as N-linked glycoprotein analogues, as α , α -disubstituted amino acids with the ability to control the secondary structure of short peptides,³ and for the introduction of carbohydrate recognition sites in synthetic peptides. This paper describes the synthesis and chemical stability of tri- and tetra-peptides which have as one of their components an amino acid at the anomeric position of protected and unprotected mannofuranose derivatives.

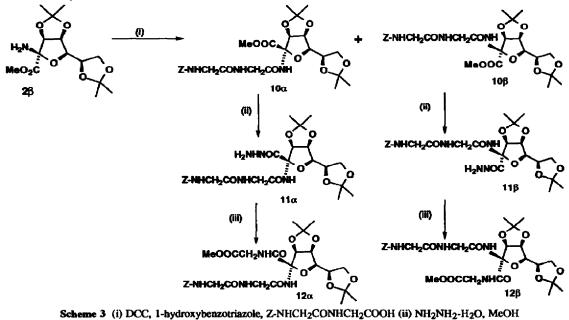


The individual epimeric amino esters 2α and 2β in dimethylformamide were separately coupled with benzyloxycarbonylglycine [ZGlyOH] activated by treatment with dicyclohexylcarbodiimide [DCC] and 1hydroxybenzotriazole to give an epimeric mixture of the protected dipeptides $5Z\alpha$ and $5Z\beta$ in 75% to 80% overall yield and a ratio of approximately 10:1 [Scheme 2]. The anomeric configuration of $5Z\alpha$ and $5Z\beta$ was determined by measurements of inter-proton distances from quantitative analysis of 500 MHz NMR NOE data.⁴ In particular, the mannose NH..H2 distance is diagnostic having possible values of 2.3 to 3.7 Å for *cis* H2/NH and 3.6 to 4.3 Å for *trans* H2/NH; from NMR data, $5Z\alpha$ has a NH..H2 distance of 2.4 Å and $5Z\beta$ a distance of 3.8 Å. In addition $5Z\alpha$ gives NH-H3 and NH-H4 NOEs, whereas $5Z\beta$ gives an NOE between NH and one of the ketal methyl groups. This is consistent only with $5Z\alpha$ having a cis H2/NH configuration.

Since the ratio of 5Z was the same regardless of which epimer of 2 was used, the rate of equilibration of the amino esters 2 was much faster than their rate of coupling; this is in marked contrast to the reactions in Scheme 1 where the reaction of the amines 2 with phenyl isocyanate was much faster than their equilibration. The thermodynamically less stable - and less hindered amine - 2α reacts significantly faster with the activated glycine ester. There was no equilibration of the dipeptides 5Z under the reaction conditions; the anomeric centre of N-acylated derivatives of 2 appears to be configurationally secure under a wide variety of acid and base conditions. The side chain acetonide in 5Z α was removed by acetic acid in methanol to give the diol 6Z,⁵ m.p. 228-229°C, $[\alpha]_D^{20}$ +92.5 (c 0.8, MeOH), in 80% yield; the diol 6Z could be converted back into 5Z α in quantitative yield by reaction with 2,2-dimethoxypropane in acetone in the presence of tosic acid.

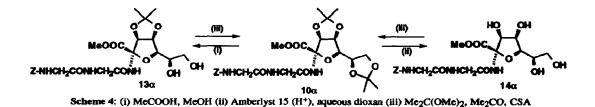
The amines 2 were also coupled with BOC-gly-OH under the same conditions to again give the α anomer 5BOC α as the major product (75% yield) with a small amount of 5BOC β (7% yield). Mild acid hydrolysis of 5BOC α gave the diol 6BOC, m.p. 224-225°C, $[\alpha]_D^{20}$ +98.5 (c 1.0, MeOH); further acid hydrolysis removed the *tert*-butyl ester rather the second ketal protecting group. No equilibration of the anomeric position of the N-acylated mannofuranose derivatives occurred during any of these transformations.

Further coupling through the C-terminus of the dipeptide $SZ\alpha$ was undertaken to demonstrate that amino acids such as 2 could be incorporated into the middle of a peptide sequence. Reaction of $SZ\alpha$ with hydrazine hydrate in methanol gave the hydrazide 7Z as an unstable white solid, $[\alpha]_D^{20} + 49.4$ (c 0.5, CHCl₃), in 87% yield. Treatment of a solution of 7Z in dimethylformamide with hydrogen chloride in dioxan, followed by *tert*-butyl nitrite,⁶ gave the acyl azide which was reacted *in situ* with glycine methyl ester hydrochloride and triethylamine to afford the protected tripeptide 8Z, m.p. 227-228°C, $[\alpha]_D^{20} + 62.0$ (c 1.0, MeOH), in 80% yield. Similar coupling of 5BOC α , via the unstable hydrazide 7BOC, $[\alpha]_D^{20} + 62.0$ (c 1.0, MeOH), gave the BOC-protected tripeptide 8BOC, m.p. 80-90(dec)°C, $[\alpha]_D^{20} + 52.4$ (c 0.5, CHCl₃), in 70% overall yield.



(iii) tert-BuONO, HCi; then Cl⁻ H₃N+CH₂COOMe, Et₃N

The aminoester 2β was also coupled with ZGlyGlyOH activated by DCC and 1-hydroxybenzotriazole to afford as the major product 10α , m.p. 99-100°C, $[\alpha]_D^{20}$ +55.0 (c 0.66, MeCN), in 74% yield, together with 10β , an oil, $[\alpha]_D^{20}$ -10.8 (c 0.98, CHCl₃) in 17% yield [Scheme 3]. The anomeric amine 2α gave 10α in 79% yield, together with 10β in 7% yield. Extension of the tripeptide 10α from the C-terminal end was effected by initial treatment with hydrazine hydrate to give the unstable hydrazide 11α , oil, $[\alpha]_D^{20}$ +46.8 (c, 1.55 CH₃CN) (68% yield); subsequent conversion to the acyl azide followed by reaction with glycine methyl ester gave the tetrapeptide 12α , $[\alpha]_D^{20}$ +57.6 (c 0.87, MeCN), in 80% yield. Similar transformations on 10β gave the epimeric tetrapeptide 12β , $[\alpha]_D^{20}$ -31.3 (c 1.08, CHCl₃), in 51% overall yield. No evidence of any epimerisation at the anomeric centre was found during any transformation after the initial acylation of the aminoesters 2.



The stability of the furanose ring under hydrolytic conditions was studied to determine whether completely unprotected sugar analogues could be readily incorporated into peptide chains. The side chain acetonide in 10 α was selectively removed by treatment with acetic acid: methanol (1:1) to give the diol 13 α , colourless oil, $[\alpha]_D^{20} + 55.6^{\circ}$ (c, 1.04 CHCl₃) in 94% yield [Scheme 4]. Reaction of 10 α with Amberlyst 15 (H⁺) ion exchange resin in dioxan:water (1:1) at 60°C removed both ketals to afford the tripeptide 14 α ,⁷ m.p. >300°C, $[\alpha]_D^{20} + 53.0$ (c 0.82, H₂O) in 78% yield. Both 13 α and 14 α could be converted back to 10 α on treatment with dimethoxypropane in acetone in the presence of camphor sulphonic acid (CSA) in 96% and 66% yields respectively. The lack of formation of any 10 β under these conditions indicated there was neither epimerisation at the anomeric centre nor interconversion of the furanose to a pyranose form during any of the hydrolysis or reprotection transformations.

In summary this paper reports for the first time the incorporation of an amino acid functionality at the anomeric position of a sugar into an oligopeptide; such materials appear to be configurationally stable at the anomeric centre to acid hydrolysis conditions, and there is no evidence of equilibration of furanose and pyranose forms. The formation of a spiro-diketopiperazine derived from $5Z\alpha$ is described in the accompanying paper.^{8, 9}

REFERENCES

⁵All new compounds in this paper have spectral data consistent with the structures proposed. Satisfactory microanalytical data has been obtained for 6Z, 6BOC, 8Z, 8BOC, 10 α , 10 β , 11 α , 12 α , 13 α ; it has not been possible to remove the last traces of DCCurea from 5 α and 5 β .

⁶Honzl, J., Rudinger, J., Coll. Czech. Chem. Commun., 1991, 26, 2333.

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¹Burton, J. W., Son, J. C., Fairbanks, A. J., Choi, S.-S., Taylor, H., Watkin, D. J., Winchester, B. G., Fleet, G. W. J., *Tetrahedron Lett.*, 1993, 34, 6199.

²Mizukai, K., Mio, S., Kawakubo, K., Honma, T., Shindo, M., Chem. Abs., 114, 81826 (1991); S. Mirza, Chem. Abs., 114, 8356 (1991).

³Deloisy, S., Thang, T. T., Olesker, A., Lukacs, G., *Tetrahedron Lett*, 1994, 35, 4783; Fauchere, J.-L., Elements for the Rational Design of Drugs in *Adv. Drgu Res.*, 1986, 15, 29.

⁴Ardron, H., Butters, T. D., Platt, F. M., Wormald, M. R., Dwek, R. A., Fleet, G. W. J., Jacob, G. S., *Tetrahedron Asymm.*, 1993, 4, 2011, full NMR and conformational analysis of these and other anomers will be published elsewhere.

⁷Selected data for 14a; $\delta_{\rm H}$ (D₂O, 200 MHz) 7.22 (5H, bs, H-Ph), 4.94 (2H, s, CH₂-Z), 4.30-4.21 (2H, m), 3.87-3.61 (7H, m), 3.55 (3H, s, CH₃-O), 3.51-3.40 (1H, m); $\delta_{\rm C}$ (D₂O): 128.8, 128.5, 127.7 (3xd, CH-Ph), 91.1 (s,C-2), 79.3, 77.4, 70.1, 68.8 (4xd, C-3,4,5,6), 67.2 (t, C-7), 62.7 (t,CH₂-Z), 53.2 (q, CH₃-O), 43.8, 41.8(2xt, CH₂-Gly); m/z (FAB+) 524 (M+K⁺, 15%), 508 (M+Na⁺, 22%), 486 (M+H⁺, 7%), 193 (100%)

⁸Estevez, J. C., Ardron, H., Wormald, M. R., Brown, D., Fleet, G. W. J., Tetrahedron Lett., 1994, 35, following paper.

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